# Study of the impact of the R693X mutation in ASXL1 on cell lines of JAK2 V617F positive myeloid neoplasias 

RAFAELA REIS VIEIRA ${ }^{1}$; BARBARA MONTE MOR ${ }^{1}$; CRISTIANA SOLZA ${ }^{2}$; ADELMO DANTAS ${ }^{3}$; DIEGO COUTINHO ${ }^{1}$; MARTIN HERNÁN BONAMINO ${ }^{1}$; ILANA ZALCBERG ${ }^{1}$<br>1. INCA, RIO DE JANEIRO - RJ - BRAZIL; 2. UERJ, RIO DE JANEIRO - RJ - BRAZIL; 3. UFF, RIO DE JANEIRO - RJ - BRAZIL.

## INTRODUCTION

The ASXL1 gene is the human homolog of Drosophila melanogaster's additional sex comb (Asx) gene, located in the region of chromosome 20q11.21, formed by 13 exons and expressed in most hematopoietic cells. The ASXL1 protein is involved in the epigenetic regulation of gene expression, interacting with histone remodeling complexes. Mutations and deletions in ASXL1 were reported in myelodysplastic syndromes (MDS) and myeloproliferative neoplasms (MPN), including chronic myelogenous leukemia (CML) and primary myelofibrosis (MF). MF is a hematopoietic stem cell clonal disorder characterized by gain-of-function mutations in JAK2, CALR or MPL genes, leading to constitutively activated JAKSTAT pathway. ASXL1 mutations can be found in about $40 \%$ of MF patients and can occur in patients with or without JAK2/CALR/MPL mutations. ASXL1 mutations were associated with an unfavorable prognosis and a higher rate of leukemic transformation. An in vitro study using cell lines has shown that homozygous ASXL1 mutations lead to the absence of protein expression (loss-offunction mutations). However, the mechanisms by which ASXL1 mutations would lead to the transformation of myeloid cells are still not fully understood. In this context, the technology of the CRISPR/Cas9 system for genetic modification of cell lines represents an interesting strategy for the study of somatic alterations of myeloid neoplasia in vitro. Here we intend to use the CRISPR/Cas9 system to insert the ASXL1 R963X mutation in the NMP cell line UKE1 and study how this molecular alteration impacts cells carrying JAK2 V617F mutation. We intend to study the methylation profile and to evaluate the protein expression of ASXL1 in these cell lines carrying mutations.


Figure 1: Representation of CRISPR/Cas9 (Savić and Schwank, 2016). In the NHEJ pathway, the ends are processed by endogenous DNA repair machinery and rejoined, which can result in random indel mutations at the site of junction and can result in gene knockout. In the HDR pathway, a repair template can be supplied which allows precise editing.

## OBJECTIVES

To evaluate the impact of the R693X mutation in ASXL1 on cell lines of JAK2 V617F positive myeloid neoplasms.

## METHODS AND RESULTS

Five different specific RNAs guides (sgRNA) for the region of interest were designed using the http://crispr.mit.edu tool and two complementary oligos for each sgRNA were designed. The vector was digested with Bbsl endonuclease linearizing vector PX458 and after we used T4 ligase to annealed oligo duplex into a specific cloning site of vector. After transformation into competent bacteria and plasmid preparation the construct was checked by sequencing using universal primer for U6 promoter region and it was possible to obtain cloned plasmids for
all five sgRNAs. First, diploid clone of the UKE-1 lineage was selected through a serial dilution of the pool and the karyotype is currently being confirmed. UKE-1 cells were currently being transfected with the plasmids obtained. After standardization we will be able to establish the study model.


Figure 2: Vector PX458 expressing Cas9 showing the cut regions of the enzyme Bbsl and specific cloning site (gRNA scaffold).


Figure 3: The five specific RNAs guides (sgRNAs) selected for the region in the ASXL1 gene where the R693X mutation (c.2077C> T, p.R693X) is found.


Figure 4: Electropherogram obtained after direct sequencing of plasmid PX458 using primer for $\mathbf{U 6}$ promoter forward region showing the guide \#1_Top annealed into a specific cloning site (gRNA scaffold).


