

Characterization of BRCA1 SQ-cluster phosphorylation status in its interaction with PALB2 in the DNA damage response

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INTRODUCTION

Germline mutations in the *BRCA1* tumor suppressor gene are associated with increased risk of hereditary breast and ovarian cancers. Its encoded protein plays a pivotal role in the maintenance of genomic integrity through homologous recombination (HR)-mediated DNA damage repair. *BRCA1* works as a scaffold to promote the formation of different protein complexes participating in several steps of HR pathway. A particular complex, formed by different cancer susceptibility gene products, including *PALB2* and *BRCA2*, plays a crucial role regulating *RAD51* recombinase activity, the crucial step of HR pathway. Mechanistically, *BRCA1* and *PALB2* interact through the heterodimerization of their coiled-coil motifs and mediate the recruitment of *BRCA2* and consequently *RAD51* to the DNA damaged sites. Recently, the association between *BRCA1* and *PALB2* was shown to be orchestrated in a cell cycle and DNA damage-dependent manner by post-translational modifications in *PALB2*, such as ATR and CDKs mediated phosphorylations. Interestingly, ATR and CDKs phosphorylation sites predicted in *BRCA1* are linked to the *PALB2*-interaction region. Those sites are associated with DNA damage response (DDR), but it is not clear whether they modulate *BRCA1/PALB2* interaction.

OBJECTIVES

This work aims to evaluate the role of *BRCA1* phosphorylation in mediating *PALB2* heterodimerization associated to DDR.

METHODS AND RESULTS

BRCA1/PALB2 interaction will be evaluated by mammalian two-hybrid and GST-pulldown assays using phosphomimetic variants generated by site-directed mutagenesis strategies - serine residues will be substituted by glutamic acids or alanine residues (mimicking a phosphorylated or unphosphorylated motif, respectively). The DDR status will be accessed by HR-proficiency assays and sensitivity to ionizing radiation/PARP inhibitors in cells expressing the phosphomimetic *BRCA1* variants. Literature curation led to identification of 8 predicted serine residues target by ATR/CDKs in the *PALB2*-interaction region of *BRCA1* protein structure. Plasmidial constructs are been generated and pilot assays testing *BRCA1/PALB2* interaction are been performed.

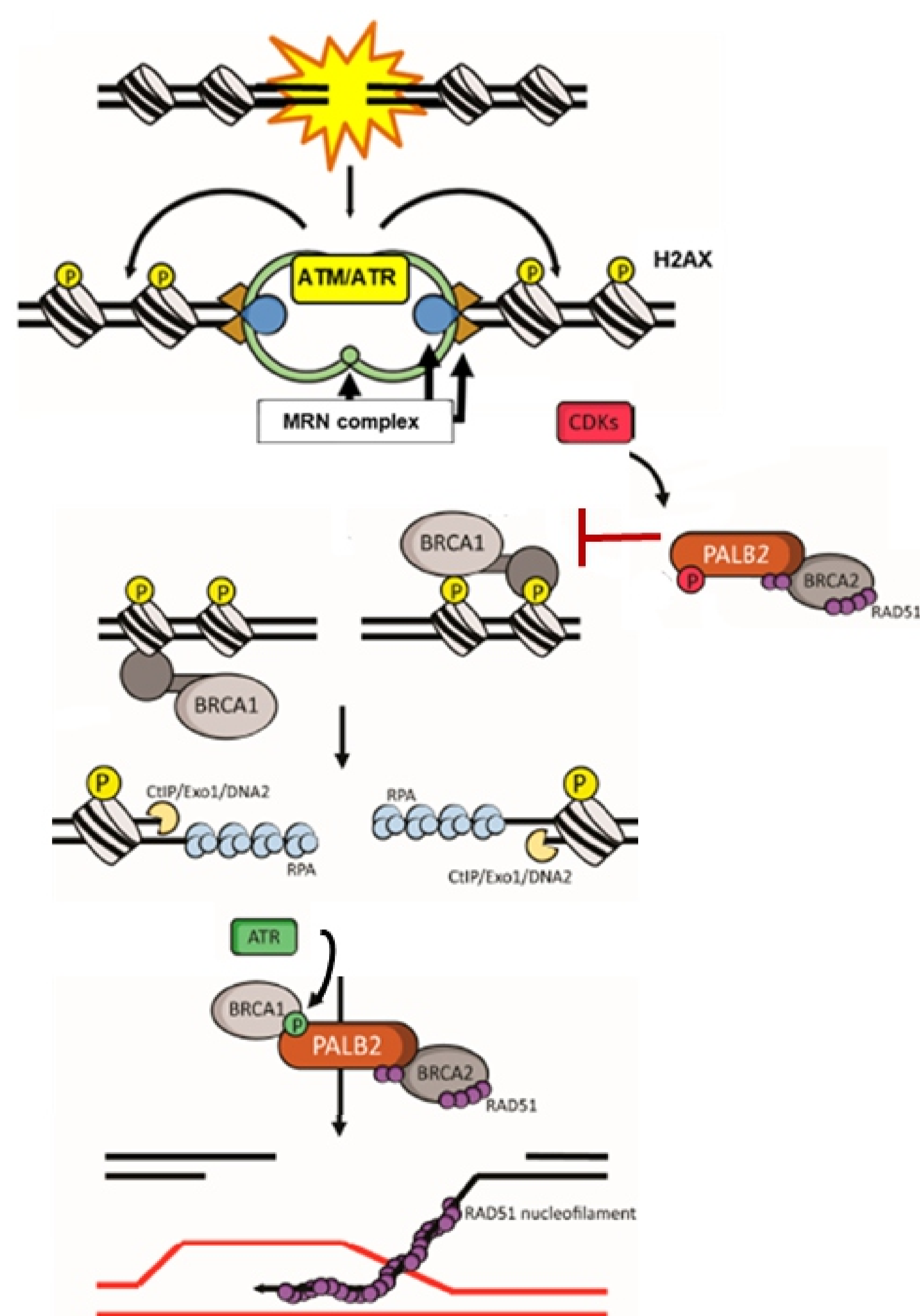


Figure 1: The phospho-modulation of *BRCA1/PALB2* association in homologous recombination (HR)-mediated DNA repair. Schematic representation of HR pathway. DNA double strand breaks are recognized by MRN sensor complex (MRE11-RAD50-NBS1) leading to ATM/ATR activation and H2AX phosphorylation (γH2AX), which in turn, amplifies the DNA-damaged signal. This phenomenon works as a platform to downstream events leading to *BRCA1* recruitment and 5' DNA end resection process (performed by CtIP, EXO1, DNA2 and MRN complex nucleases activities stimulated by *BRCA1*). A CDK-mediated *PALB2* phosphorylation abrogates its association with *BRCA1* at this moment. Afterwards, RPA caps exposed single-strand DNA and are substituted by *RAD51* recombinase in a process dependent on *BRCA1/PALB2/BRCA2* complex. At this point, a ATR-mediated *PALB2* phosphorylation promotes its association with *BRCA1*, and consequently, *RAD51* recruitment and DNA damage resolution.

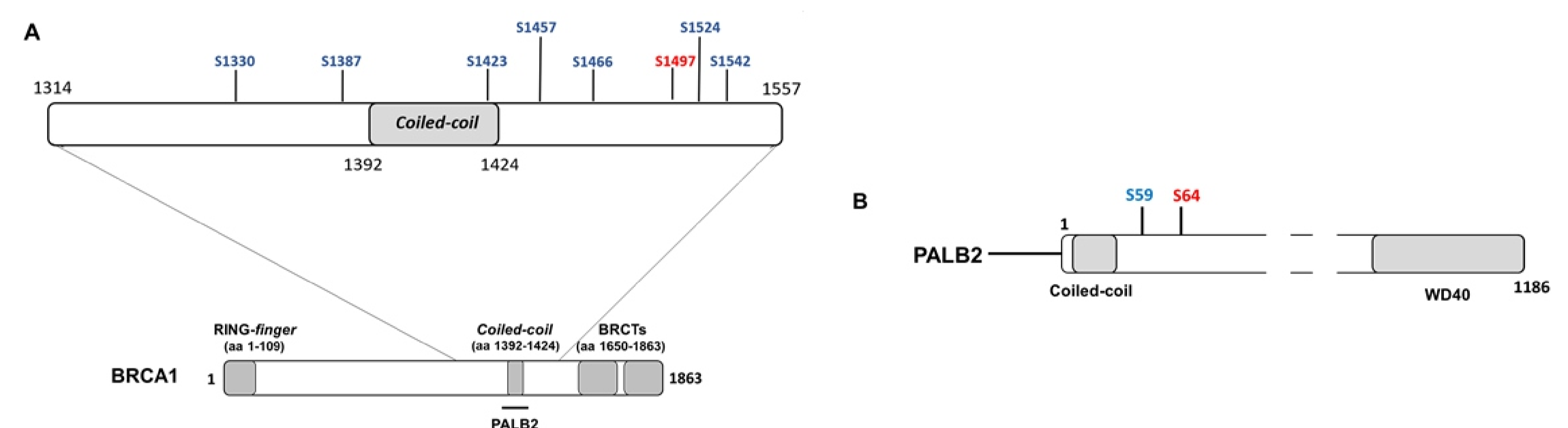


Figure 2: Phospho-modulation of *BRCA1/PALB2* association. (A) Schematic representation of *BRCA1* protein with the SQ-cluster depicted and (B) *PALB2* protein. CDKs phosphosites are highlighted in red and ATM/ATR in blue. Protein domains are represented in gray.

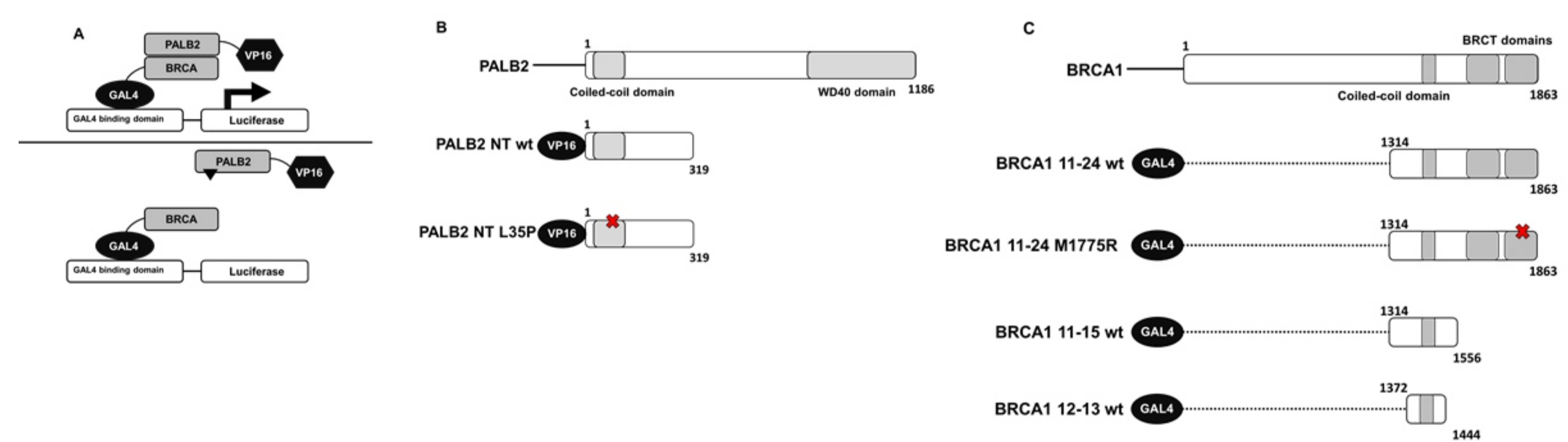


Figure 3: Mammalian two-hybrid (M2H) assay. (A) Schematic representation of M2H assay. (B) Schematic representation of *PALB2* N-terminal (NT) and (C) *BRCA1* constructs.

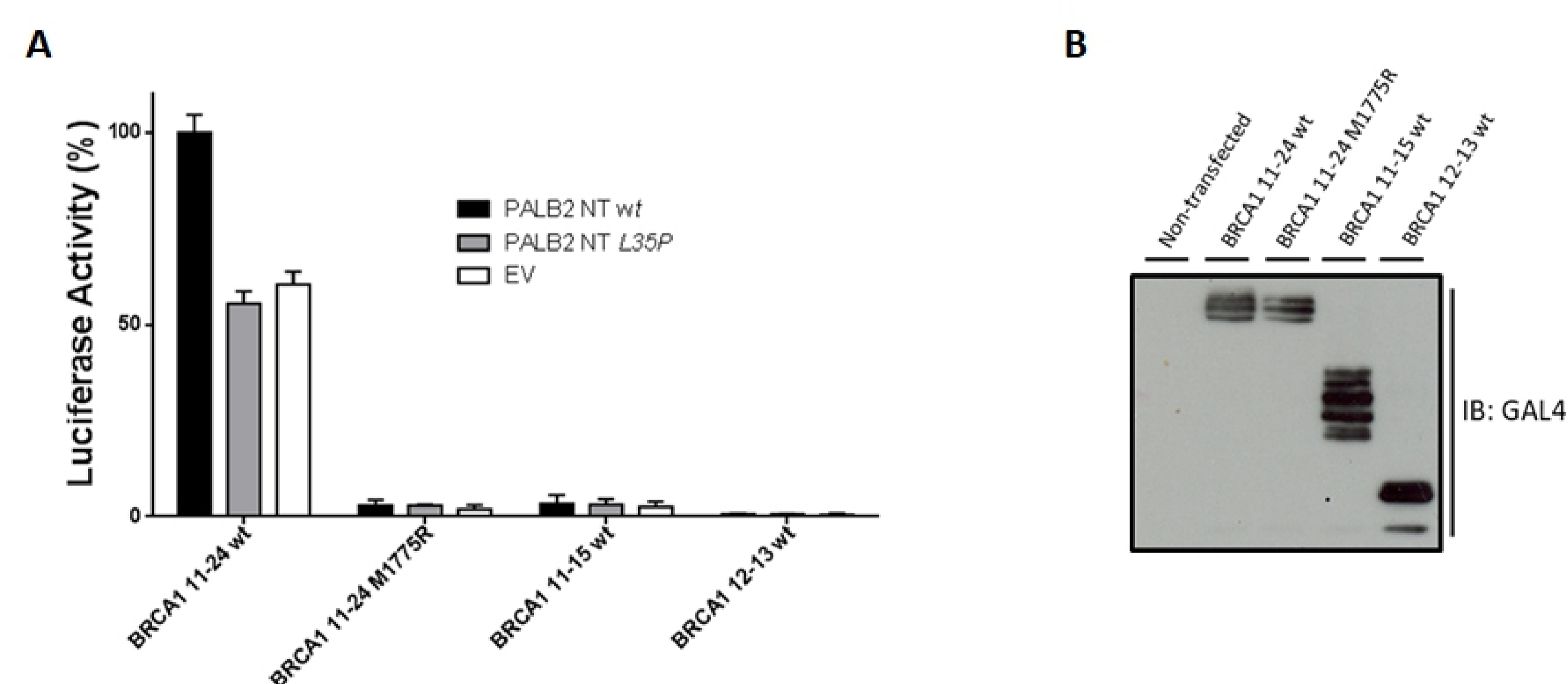


Figure 4: Mammalian two-hybrid assay. (A) HEK293FT cells were cotransfected with the reporters (pG5/*uc* and pGR-TK), *BRCA1* constructs (*BRCA1* 11-24 wt, *BRCA1* 11-24 M1775R, *BRCA1* 11-15 or *BRCA1* 12-13) and *PALB2* coding plasmids (wt, L35P or empty vector (EV)); the mammalian two-hybrid assay was performed 24h post-transfection. Data represent luciferase activity of a representative experiment of a series of independent experiments. (B) Immunoblotting analysis of M2H proteins.

CONCLUSIONS

The comprehension of the molecular mechanisms behind *BRCA1/PALB2* association is extremely important for a better understanding of maintenance of genomic integrity and cancer predisposition. It also impacts in patient surveillance and in the development of new treatment based on *BRCA*'s status.