

# PGE<sub>2</sub> induces epithelium-mesenchymal transition in colorectal cancer cells

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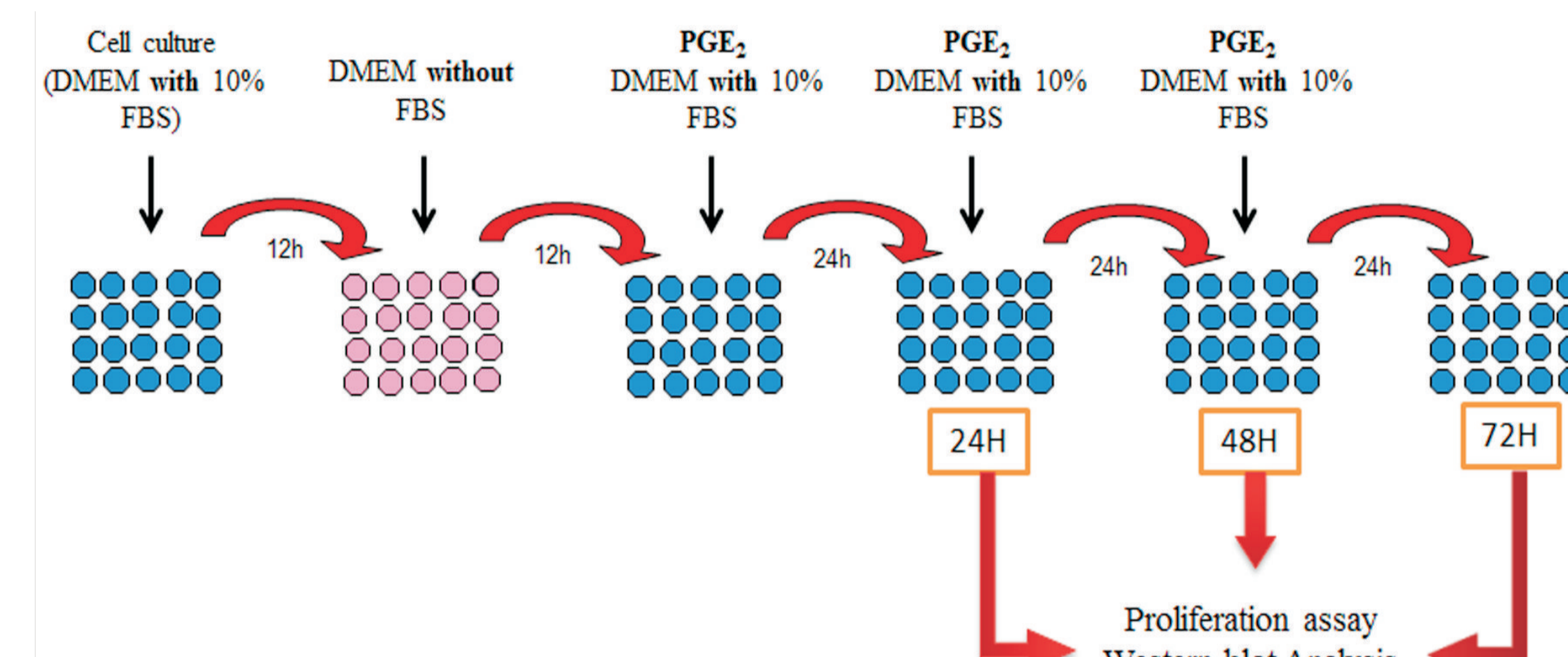
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## ABSTRACT

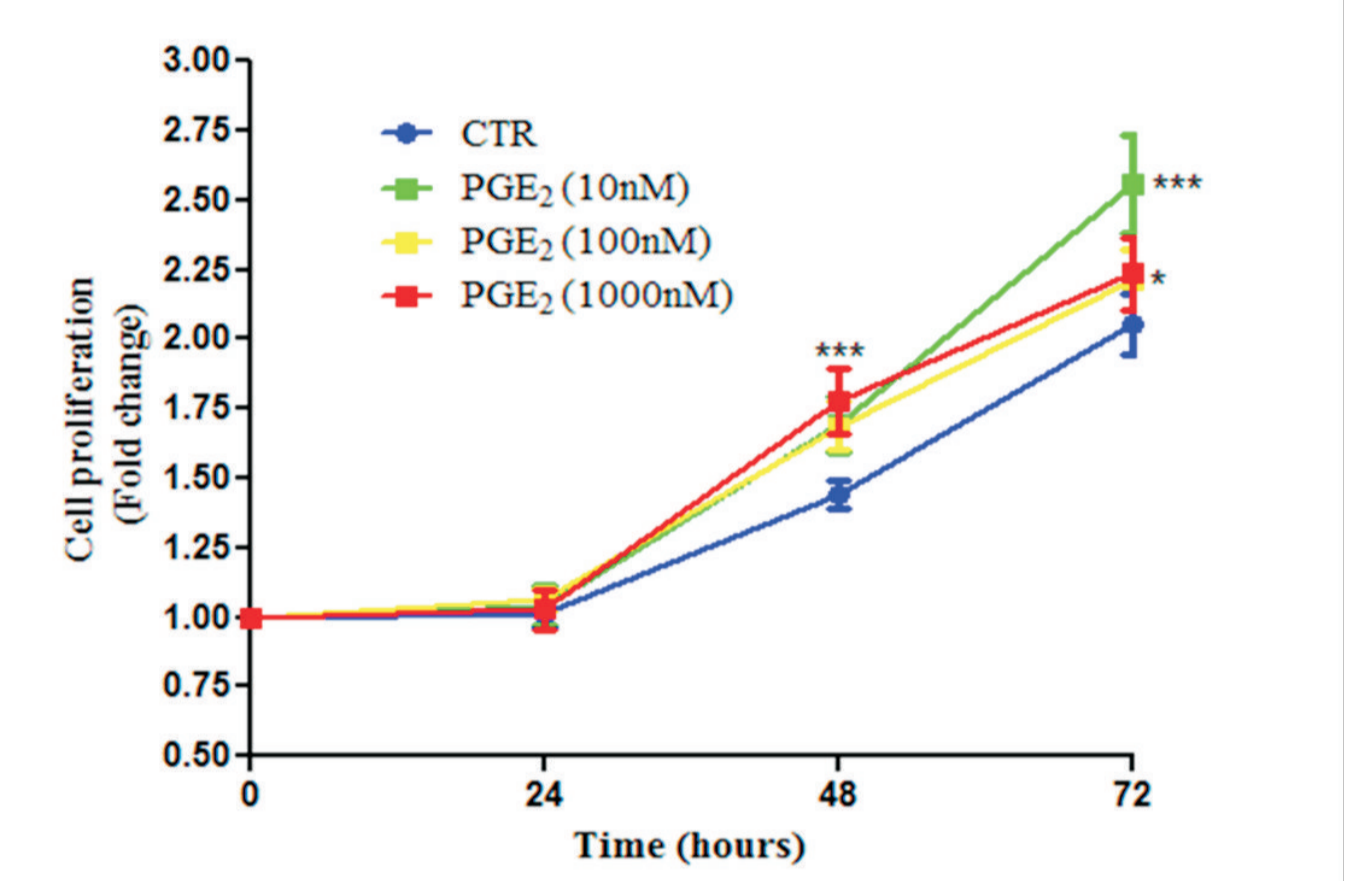
**Background:** The inflammatory process is one of the most important risk factors in colorectal cancer (CRC) and the increased expression of COX-2 enzyme is associated with this process. It is known that prostaglandin E<sub>2</sub> (PGE<sub>2</sub>), the main product of COX<sub>2</sub>, is a potent inducer of tumor progression, increasing the metastasis ability of cancer cells. During initial steps of cancer progression the cells acquire an undifferentiated phenotype providing a more migratory and invasive potential, an event known as epithelial-mesenchymal transition (EMT). In this context, the role that PGE<sub>2</sub> plays during the EMT is not completely understood. Aims: To analyse the role of PGE<sub>2</sub> during the EMT development in CRC.

**Methods:** CRC cells, HT-29, were treated with different concentrations of PGE<sub>2</sub> and the proliferation rates were analyzed after 24, 48 and 72h. Western blotting (WB) and immunofluorescence (IF) assays were employed to analyze the expression and subcellular localization of epithelial and mesenchymal markers. Additionally, analysis by polymerase chain reaction was conducted to evaluate the PGE<sub>2</sub> receptors profile.

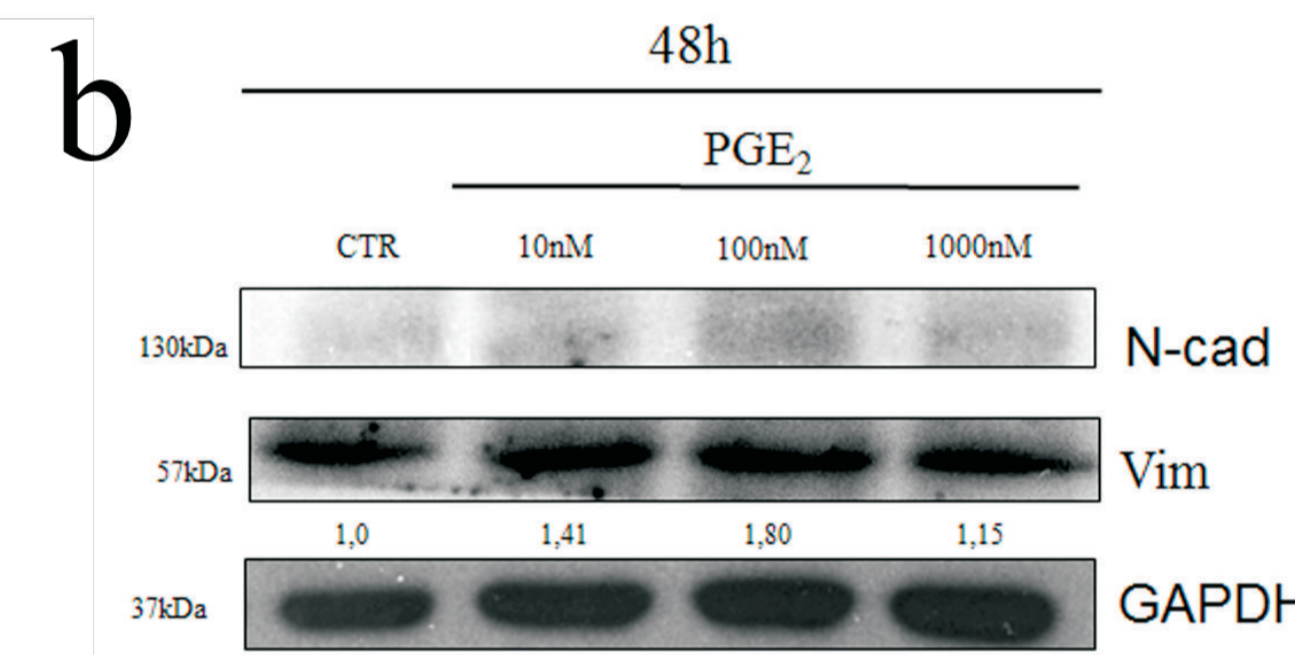
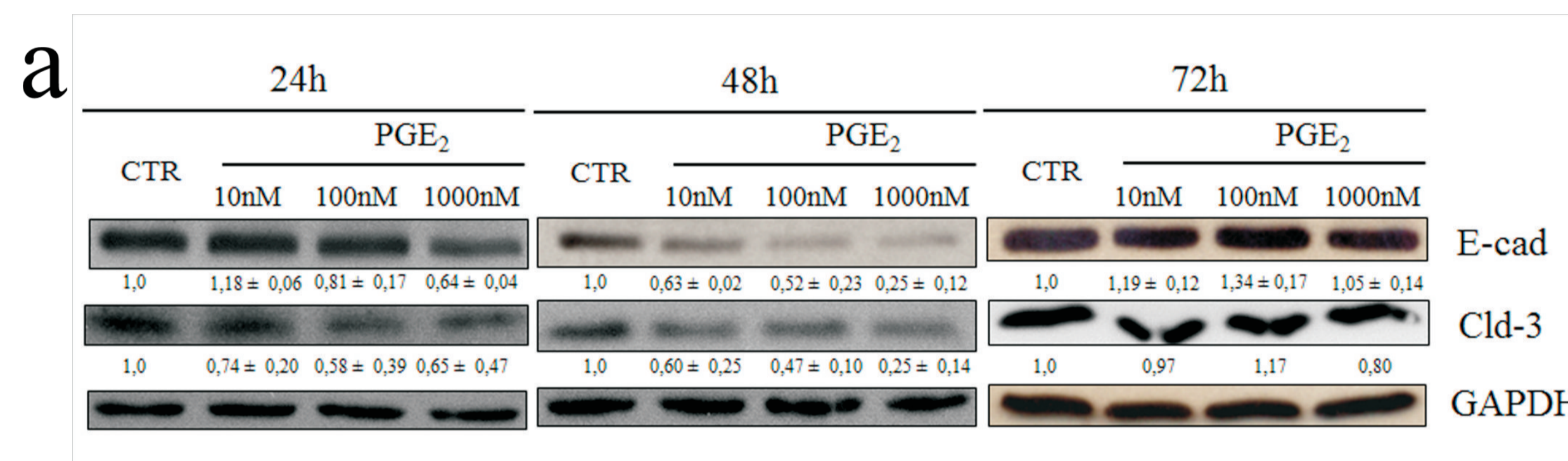
**Results:** 1000nM PGE<sub>2</sub> increased the proliferation of HT-29 cells after 48h of treatment. We observed that PGE<sub>2</sub> in this time and concentration induced a decrease of E-cadherin and claudin 3 expression as well as protein redistribution from the cell-cell contacts. In addition, IF for beta-catenin suggests translocation of the protein to the nucleus in the treated cells. Conclusion: HT-29 cell treated with PGE<sub>2</sub> display events related with EMT development and these results are the basis to determine the mechanistic by which PGE<sub>2</sub> induces EMT.



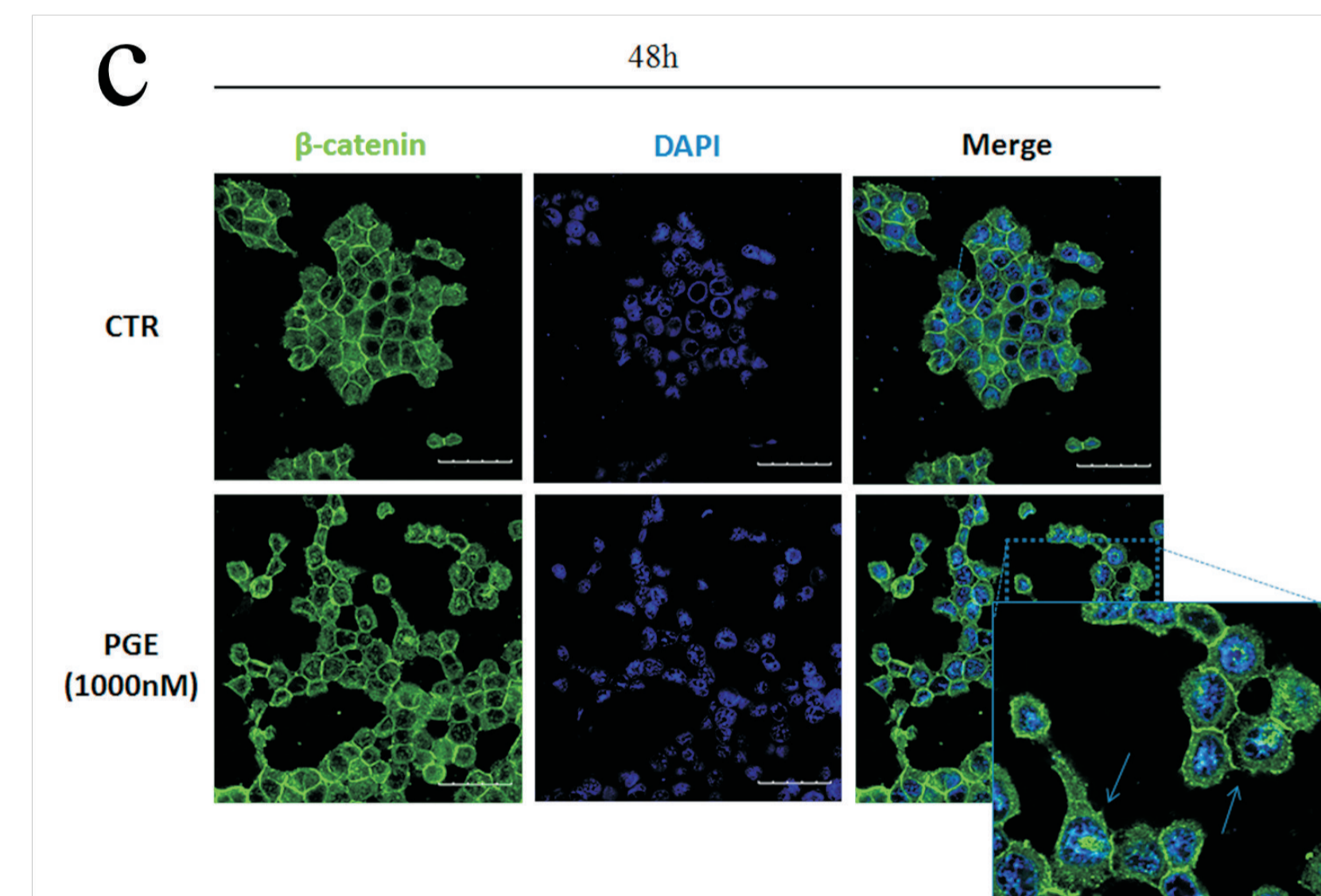
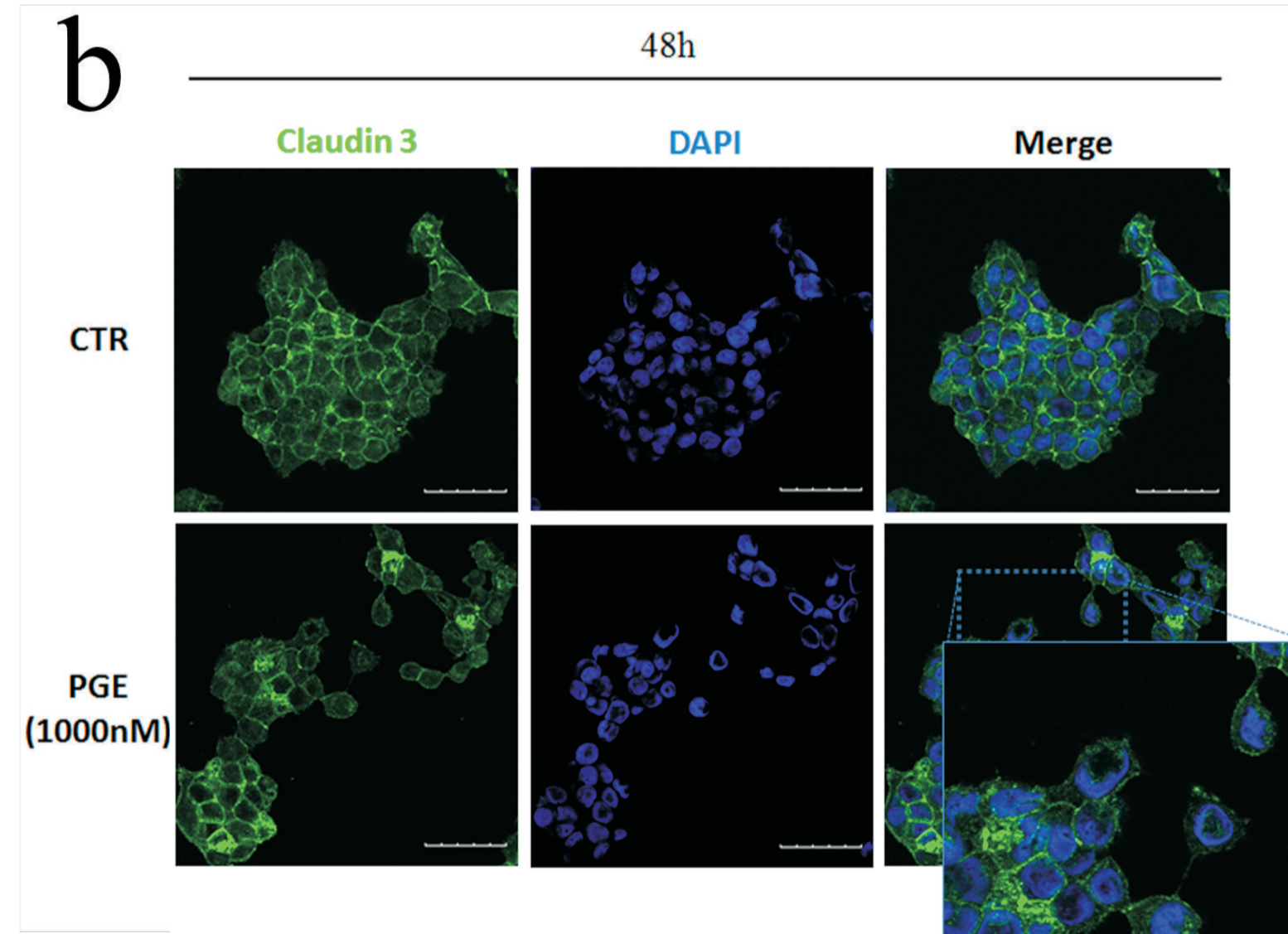
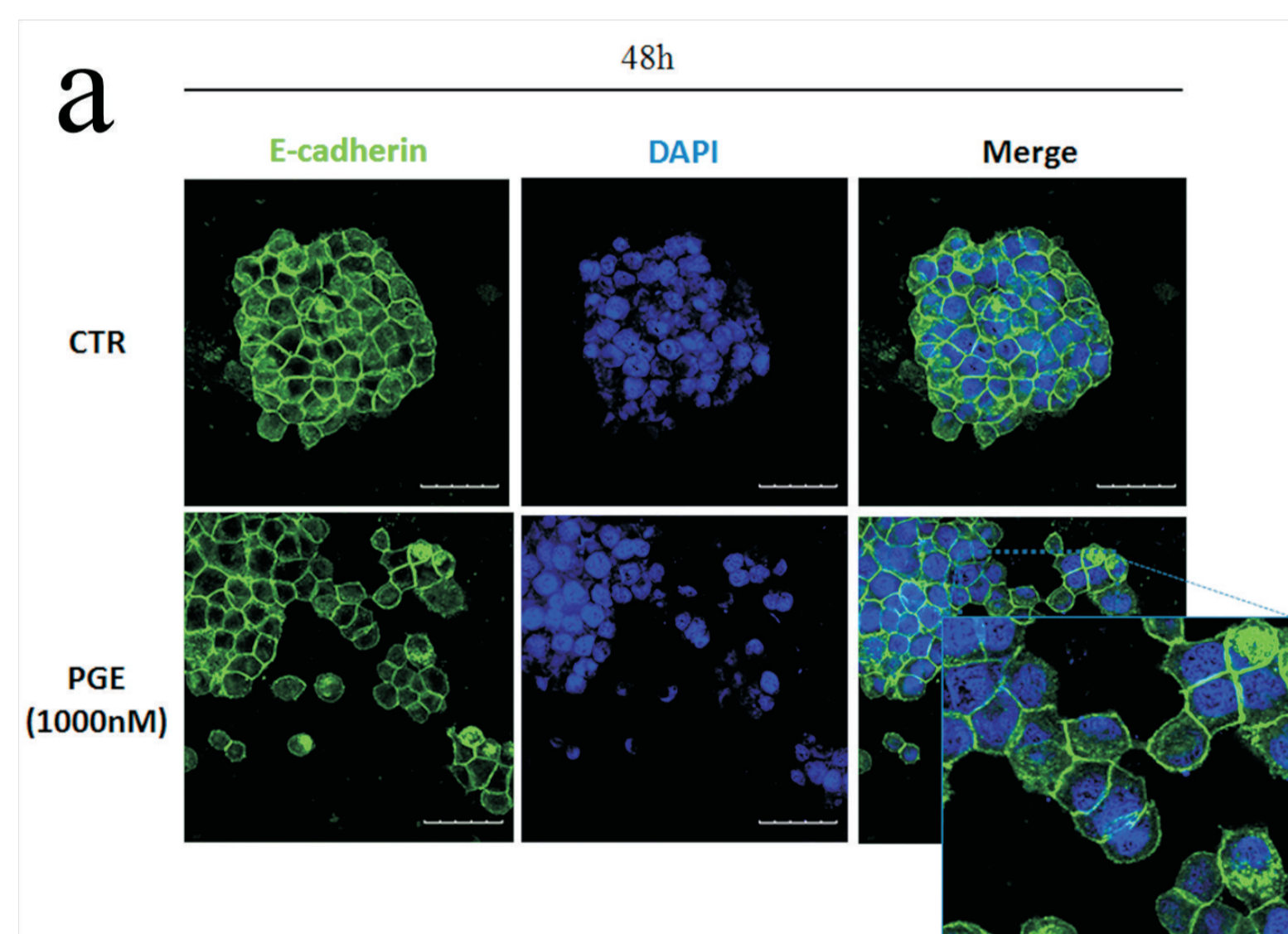
**Fig. 1.** Protocol of treatment with PGE<sub>2</sub>. Before PGE<sub>2</sub> treatment, the human colon cancer cell line (HT-29) was maintained in DMEM with 10% fetal bovine serum (FBS) during an incubation period of 12 hours and then was starved in serum-free medium for 24 hours. The treatment with PGE<sub>2</sub> in different concentrations (10nM, 100nM and 1000nM) was reapplied every day for each time of analyses (24, 48 and 72 hours).



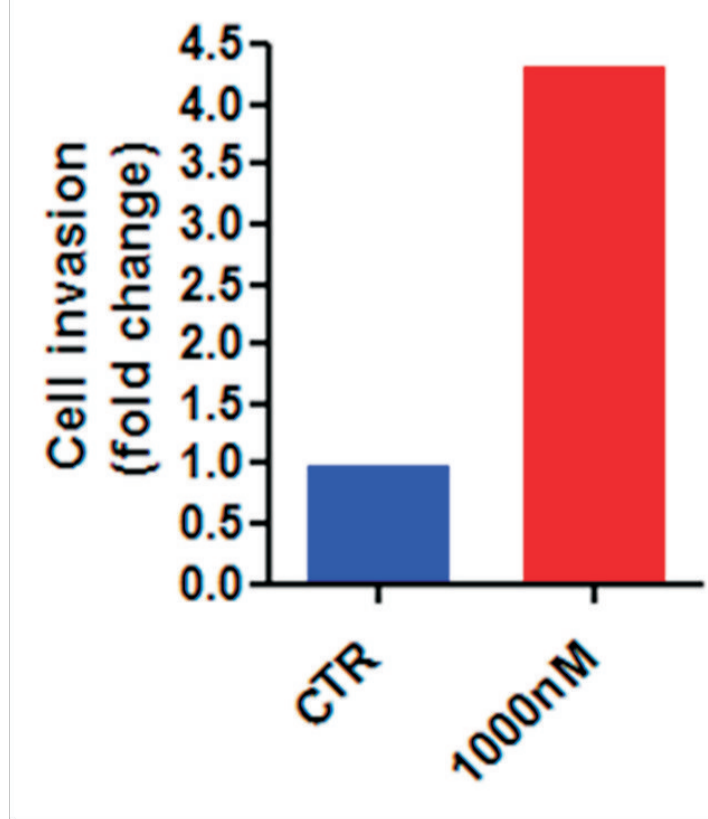
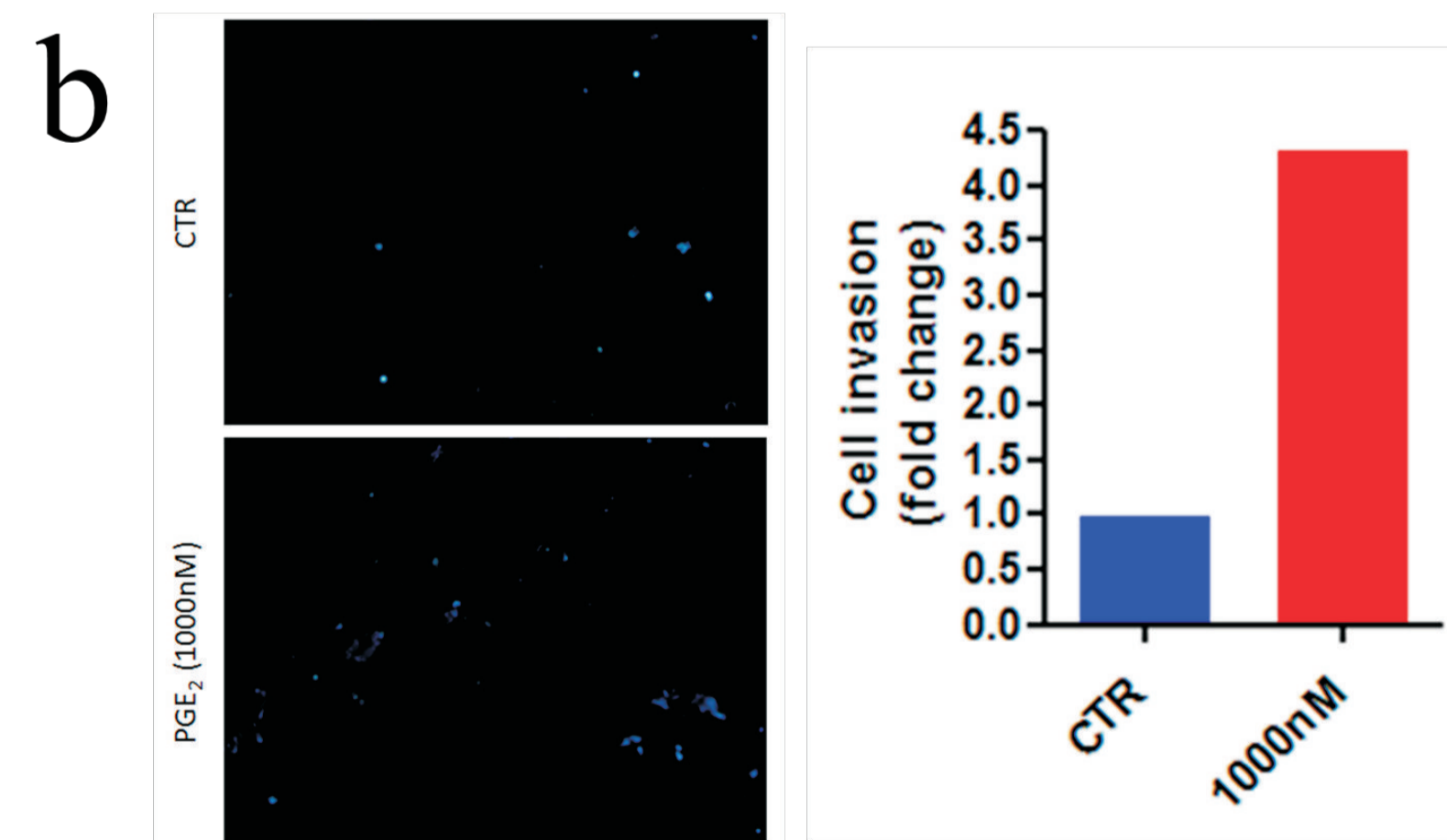
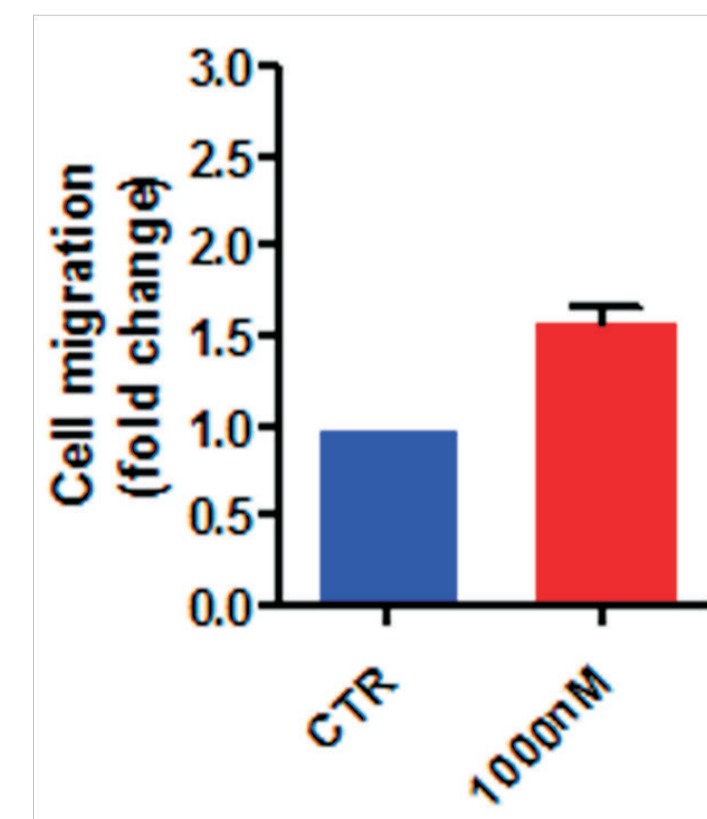
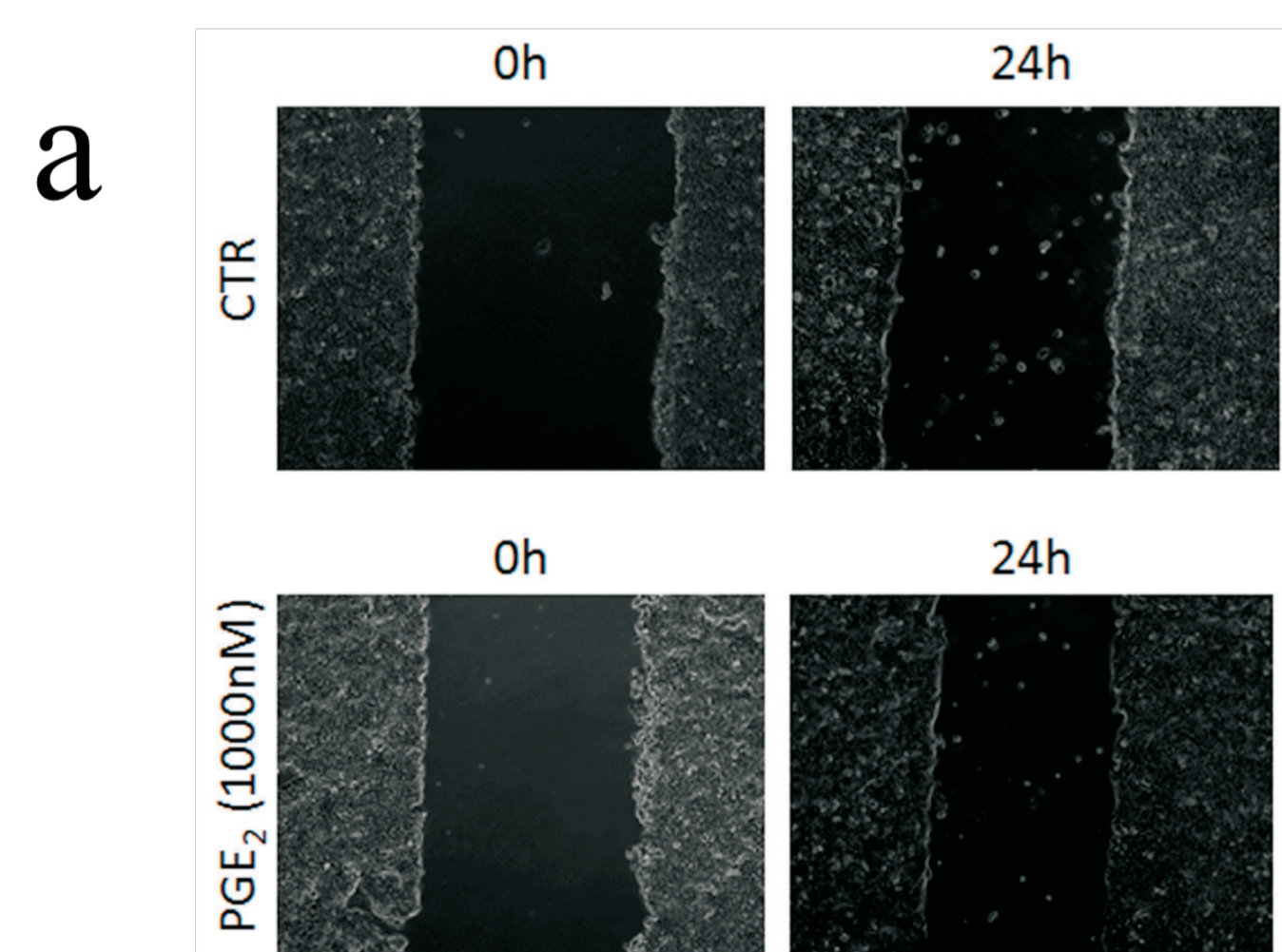
**Fig. 2.** The treatment with PGE<sub>2</sub> induces cell proliferation in HT-29 cell. HT-29 cells were treated with PGE<sub>2</sub> (10nM, 100nM and 1000nM) and the proliferative effect was analysed by violet cristal assay. The results are expressed as mean ± s.e.m. of three independent experiments. \**P*<0.05; \*\**P*<0.005; \*\*\**P*<0.001 compared with control (CTR).



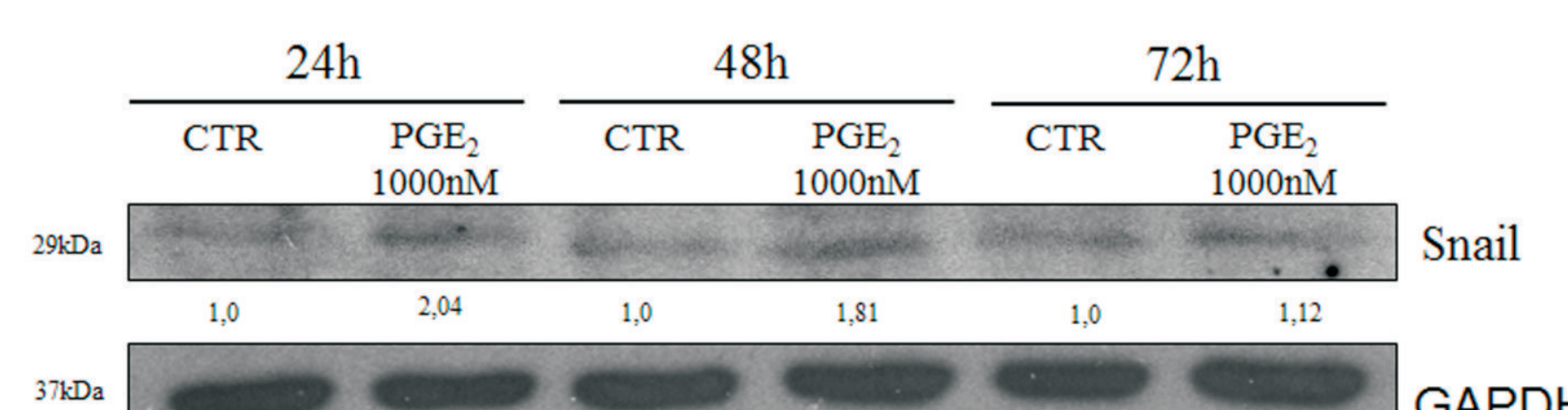
**Fig. 3.** PGE<sub>2</sub> induces decreased of epithelial proteins expression and enhanced mesenchymal proteins expression in HT-29 cell. HT-29 cells were treated with PGE<sub>2</sub> (10nM, 100nM and 1000nM) and analyzed by immunoblotting assay. a E-cadherin and claudin 3 protein levels in lysates were analyzed after 24, 48 and 72h of treatment. The results are expressed as mean ± s.e.m. of three independent experiments. b N-cadherin and vimentin protein levels in lysates were analyzed after 48h of treatment. Numbers below the figure represent the ratio of the optical density of the bands as fold change of protein expression normalised by GAPDH.



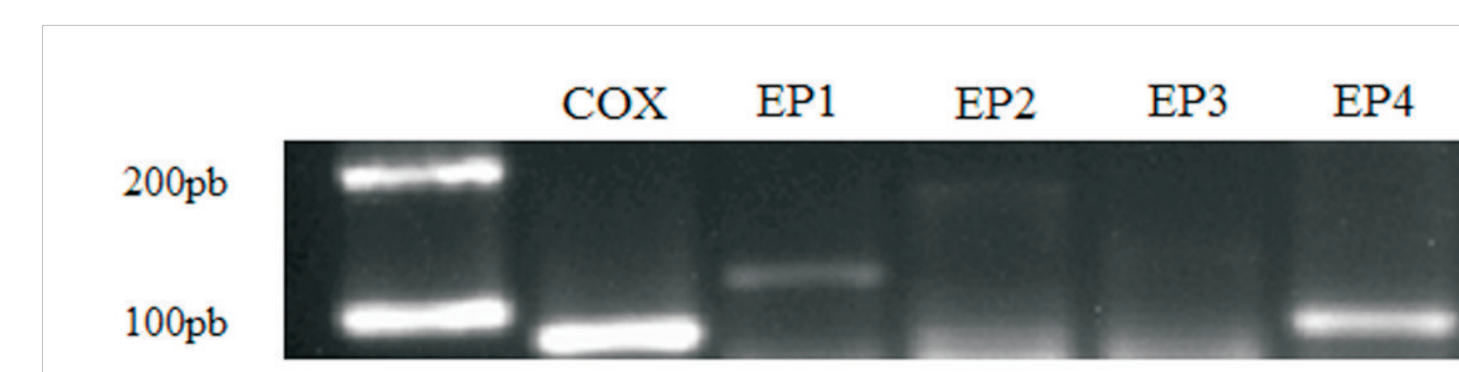
**Fig. 4.** Subcellular localization of E-cadherin and claudin 3 and nuclear translocation of beta-catenin upon 48h of treatment with 1000nM PGE<sub>2</sub> in HT-29 cell. HT-29 cells were grown in glass coverslips until colony formation and treated or not with 1000nM PGE<sub>2</sub> for 48h, after 24h of starvation. Then, cells were subjected to immunofluorescence analysis of: a E-cadherin, b Claudin 3, c beta-catenin. The nucleus are stained with DAPI. Scale bar, 50µm.



**Fig. 5.** Treatment with 1000nM PGE<sub>2</sub> enhanced HT-29 cell migration and cell invasion in 24 hours and 48h, respectively. a HT-29 cells were treated or not with 1000nM PGE<sub>2</sub> and analyzed by wound healing assay at 0h and 24h of treatment. Bar graph are plotted as a fold change of cell migration (where non-treated cells = 1). Data are presented as the mean ± s.e.m. of two independent experiments. b HT-29 cells were treated or not with 1000nM PGE<sub>2</sub> and the invasiveness was analyzed by Transwell assay at 48h of treatment. Bar graph are plotted as a fold change of cell invasion (where non-treated cells = 1).



**Fig. 6.** PGE<sub>2</sub> induces increased of the transcriptional factor Snail expression in HT-29 cell. HT-29 cells were treated with PGE<sub>2</sub> (1000nM) for 24, 48 and 72 hours and Snail protein levels in lysates were analyzed analyzed by immunoblotting assay. Numbers below the figure represent the ratio of the optical density of the bands as fold change of protein expression normalised by GAPDH.



**Fig. 7.** HT-29 cells express mRNA of three PGE<sub>2</sub> receptors (EP1, EP2 and EP4) and COX<sub>2</sub> enzyme. HT-29 cells were cultured following the protocol with starvation for 24h and after maintained just in DMEM with 10% fetal bovine serum (FBS) for 48h. Conventional PCR of mRNA for all PGE<sub>2</sub> receptors and COX<sub>2</sub> enzyme are represented in the figure.