EVALUATION OF A COMPOSITE SKIN SUBSTITUTE TO TREAT VENOUS STASIS ULCERS: A Pilot Study

Avaliação do uso de substituto de pele composto no tratamento de úlceras de estase venosa: um estudo piloto

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ABSTRACT

Introduction: Venous leg ulcers represent a therapeutic challenge, and they are associated to significant morbidity and cost. Many advances have been made to develop and apply biological skin substitutes, showing they can be beneficial for patients with burns, leg ulcers, and other skin lesions. Autologous cultivated keratinocytes have been demonstrated to be useful in leg ulcer treatment, but the lack of a dermal component may compromise their effects and the quality of the scar. **Purpose:** To evaluate a composite skin graft of human acellular dermis and autologous keratinocyte culture to treat leg ulcer. **Methods:** Two patients with four long-term chronic ulcers were treated with a composite skin graft of human acellular dermis and autologous keratinocyte culture for a month in this pilot study. **Results:** The four ulcers had a surface reduction of 72.21%, 36.36%, 60.93%, and 15.90% after one month of treatment. No adverse tissue reactions were observed. Histology and immunohistochemistry showed that humam acellular dermis was free of donor's cells, and the composite skin substitute preserved the architecture of the normal skin, including the basement membrane components and stratified epidermis. **Conclusions:** These preliminary observations suggest that such composite skin substitute could be an alternative to recalcitrant leg ulcers.

Keywords: Varicose Ulcer; Extracellular Matrix; Skin, Artificial; Tissue Culture Techniques; Cell Culture Techniques

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INTRODUCTION

Leg ulcers are quite frequent and associated to significant morbidity, high medical costs and major impact on the quality of life and work productivity. Venous stasis ulcers represent 70-90% of cases1, and their management represents a hard medical challenge. It is a consensus that compression is the main step to heal venous ulcers^{2,3}, and, therefore, it is considered the gold standard therapy for such condition⁴. However, the healing rates can be as low as 22% after 12 week therapy and the 12-month recurrence rate, as high as 69%5. Over the last years, the development of biological skin substitutes brought new strategies to treat difficult-to-heal ulcers6. Keratinocyte sheet autografts have proved to be life saving to treat large third degree burns and have been successfully employed in managing chronic ulcers7. Epidermal equivalents prepared from autologous hair follicle keratinocytes applied on recurrent leg ulcers led to reepithelialization of 70% of the total wound surface after 8 weeks with a healing rate of 32%.⁸ Although their advantages are permanent and with rapid wound coverage, application onto large areas with material obtained from a small skin biopsy, and faster pain relief⁹, they present some inconveniences. They are fragile to manipulate, resulting in an unstable epithelium, giving rise to spontaneous blistering many months after grafting. Additionally, they have an increased susceptibility to infections and contractures¹⁰. Histologically, the fragility of the grafted cultured epithelial autograft sheets may be related to the incomplete dermalepidermal junction, giving rise to inadequate anchoring¹⁰. In order to improve the quality of the healed skin, some authors suggest the use of dermal replacement beneath these cultured epithelial grafts.¹¹

Thus, a skin substitute constituted by a fibroblast-derived dermal replacement (Dermagraft[®]) tested in patients with venous leg ulcers showed to improve the healing rate and the peri-ulcer skin perfusion¹². A series of patients in a multicenter, prospective, randomized, controlled trial showed that the use of an allogeneic bilayered cultured skin equivalent (Apligraft[®]) on hard-to-heal venous leg ulcers allowed a complete wound closure in 6 months¹³. It seems that Apligraft is more effective and less expensive in healing venous leg ulcers than conventional therapy alone¹⁴⁻¹⁷. Several types of simple composite substitutes have been developed, and one of them is acellular dermis covered with autologous keratinocytes¹⁸. Acellular human dermis is an allograft from which epidermal cellular components have been removed, with preservation of the dermal surface architecture and basement membrane complex. The process of removing the cellular elements, the most immunogenic components of an allograft skin, makes it an immunologic inert dermal tissue¹⁹.

Rennenkampff and colleagues examined the effects of an acellular dermal matrix on skin repair, and demonstrated that human keratinocytes persisted in all animals grafted with dermal matrix, compared to only 63.6% of animals with no dermal component ¹¹. They also showed a significantly reduced wound contraction in animals receiving dermal matrix.

Considering the therapeutic challenge represented by venous stasis ulcers, two patients with recalcitrant leg ulcers were treated with a composite skin substitute formed by human acellular dermis, and cultivated autologous keratinocytes, in order to establish the feasibility of the procedure as well as the tissue response to it.

METHODS

Patients - Two patients enrolled for the study were inpatients at the Clementino Fraga Filho University Hospital, who signed the informed consent as outlined for the project approved by the Hospital's Ethical Committee.

Case 1 - A 55 year old black man with two five-year old ulcers located at the maleolar external and internal regions of the left leg, measuring 10.15 cm^2 and 11.00 cm^2 , respectively. Clinically, there were signs of venous insufficiency, with preservation of the arterial pulses. Dopplerfluxometry was compatible with venous insufficiency.

Case 2 - A 65 year old black man with three ten-year old ulcers. Patient had a history of traumatic ulcer (burn) and he had been submitted to a skin graft with no clinical improvement in the previous years. The ulcers were located on the anterior tibial area of the right leg, and they measured 10.65 cm², 0.70cm², and 6.35 cm², respectively. The skin was covered by an atrophic scar, and there were signs of venous insufficiency. Arterial pulses were preserved and dopplerfluxometry was compatible with venous insufficiency.

In both patients, the autologous keratinocytes were obtained from a skin biopsy on the dorsum. Five and seven weeks later for case 1 and 2, respectively, the composite skin graft was applied over the ulcers. Before applying the skin substitute, the ulcers were cleaned with saline solution and superficially debridated after local anesthesia. The dressing used was AdapticTM (Johnson & Johnson). The dressing was changed 5 days after the application and every 3 days for 8 weeks afterwards.

Keratinocyte cell culture – Normal skin autologous keratinocytes were obtained according to Green and colleagues²⁰. Biopsies were cut into fragments and submitted to mechanical dissociation of the epidermis from dermis followed by enzymatic dissociation with a 0.3% trypsin in calcium and magnesium-free saline solution. Cells were harvested by centrifugation and cocultured with 3-T3-J2 mouse fibroblasts (obtained from the Rio de Janeiro Cell Bank), pretreated with 15 µg/ml mitomycin-C solution (Boehringer-Mannheim, Indianapolis, IN). They were maintained in keratinocyte culture medium consisting of a 1:1 mixture of Dulbecco's (DMEM, high glucose) and Ham's F12 media (both from GIBCO/BRL, Gaithersburg, MD, USA), supplemented with 10% fetal bovine serum (JRH Bioscience, Lenexa, KS), adenine, 1.8 x 10-4 M (Sigma Chemical, St Louis, MO, USA), cholera toxin, 10-10 M (Vibrio Cholerae, type Inaba 569 B; Calbiochem, La Jolla, CA, USA), hydrocortisone, 0.4 µg/ml (Novo Nordisk, Princeton, NJ, USA), transferrin, 5µg/ml (Boehringer-Mannheim), triiodo-l-thyronine, 2x 10-9 M (Sigma), amphotericin B, 2.5 µg/ml, and penicillin-streptomycin, 100IU/ml to 100 µg/ml (Boehringer-Mannheim), and epidermal growth factor, 10 µg/ml (Collaborative Biomedical Products, Bedford, MA, USA). Cultures were incubated in a humidified 10% CO2, at 37° C.

Preparation of the acellular dermis – Acellular dermis was prepared from donor cadaver skin obtained from Rio Transplante Skin Bank (hepatitis B- and HIV negative). Acellularization was obtained as to the modified protocol described by Ralston and colleagues²¹.

Fragments of skin were submitted to three rapid freeze-thaw cycles in liquid nitrogen to devitalize cells, washed three times in sterile PBS, and immersed in a series of increasing concentration of glycerol in phosphate saline buffers.

After being dried, acellular dermis was sterilized in ethylene oxide, rehydrated, and extensively washed in an 1 M NaCl solution. The epidermis was gently stripped from the dermis with a forceps, and the dermis was immersed in PBS with antibiotics solution (gentamycin at 100 μ g/ml, ciprofloxacin at 10 μ g/ml, amphotericin B at 2.5 μ g/ml, penicillin-streptomycin at 100 U/ml to 100 μ g/ml) at 40C for 4 weeks or more. Before the use, the acellular dermis was washed three times in a saline solution to remove residual antibiotics.

Preparation of the composite skin grafts – Pieces of acellular dermis were placed into a 35-mm tissue culture dishes, with the papillary side up. Cultured keratinocytes (0.5 ml of 1.0 x 106 cells/ml) were seeded on top of the papillary surface of the acellular dermis and maintained 24 hours immersed in DMEM supplemented with 10% fetal bovine serum. This "skin composite" was subsequently raised and maintained at the air-liquid interface for 1 week prior to grafting.

Clinical assessment – Two months of following patients up after the surgery each week. Photographs of ulcers were taken with a digital SONY Cybershot 2.1 megapixels camera, and the areas were measured by the software Image Tool for Windows (version 3.00), and a mean of three consecutive measurements was performed.

Immunostaining and histology – To monitor the structure of the composite skin substitute, a fragment was processed by routine histology techniques.

An immunohistochemical study on paraffin sections characterized the presence of laminin and collagen type IV (DAKO, Carpinteria, CA, USA). Antibodies were detected with LSAB+ - HRP (DAKO), followed by diaminobenzidyne (DAB, DAKO), and hematoxylin counterstaining.

RESULTS

Composite skin substitutes' biopsy showed a stratified epidermis with presence of laminin and type IV collagen at the dermo-epidermal junction, as well as a complete acellularized dermis maintaining its original structure (Figure 1). To the application, grafts had approximately the same size of each ulcer. After the graft application, the integration of the composite skin substitute was visually recognizable, in both patients. The graft adhered to the underlying wound within 5 days. Patients referred pain reduction. They did not present any complain and no adverse or side effects related to the application were noticeable upon clinical examination.

Case 1 – One month after the application of the skin substitute, the ulcers' diameters reduced from 10.15 cm² to 2.82 cm² and from 11.00 cm² to 7.00cm², representing a reduction rate of 72.21% and 36.36% respectively.

Case 2 – After 1 month of the composite skin substitute application, the area of the ulcers reduced from 10.65 cm^2 to 4.16 cm^2 , and from 6.35 cm^2 to 5.34 cm^2 . The reduction rate were repectively: 60.93% and 15.90%. The third and smaller ulcer (0.70 cm^2) completely healed. (Figure 2).

Figure 1: A) Human acellular dermis – Immunohistochemistry to collagen IV. Reactivity can be observed at the dermo-epidermal junction (*) and the vascular wall. Scale bar 1µm. 100x

B) Composite skin substitute. Hematoxyllin Eosin X 100. It can be observed a stratified and differentiated epidermis with formation of a corneal layer (\downarrow). Scale bar 1µm. 100x



Figure 2: A) Case 2 patient, before applying the skin substitute. B) Same patient, 34 days after the application of the skin substitute, showing a size reduction of the ulcer.





Figure 3 – A) Case 1 patient, before applying the skin substitute. B) Same patient, 43 days after the application of the skin substitute, showing a size reduction of the ulcer.

DISCUSSION

The established treatment protocols to leg ulcers, including debridement, compression therapy, and venous surgery optimized the wound healing environment but did not stimulate healing directly¹⁵. New advances in wound healing must provide an active stimulus to the wound.

Although split-thickness autograft remains an option, the pain at the donor site, healing delays, infection and hypertrophic scarring are a limitation for its current use²². The optimum characteristics of a skin substitute are: 1. rapid attachment to the wound allowing the revascularization; 2. provide a barrier against fluid loss and infection; and 3. easy handling²³. Additionally, it should be nonantigenic, with minimal inflammatory or foreign body response and nontoxic⁹.

Although autologous culture keratinocytes may be useful to treat leg ulcers and to provide a new epidermal surface, they are difficult to handle, generating an epidermis subject to physical stress, resulting in early graft loss, blistering and scarring¹⁸. The use of composite skin grafts here described disclosed no adverse events, made the manipulation of the graft easier, and increased the graft integration. Preservation of the basement membrane complex at the dermo-epidermal junction granted better mechanical properties of the healing skin. Although treated patients did not completely heal the ulcers in one month, their size had a significant reduction as well as morbidity. It is relevant to emphasize that those were long standing ulcers (5 and 10 years evolution), with larger than 5 cm² area were considered refractory to traditional treatments. Prognostic indicators of venous ulcers demonstrated that in subjects with a greater than 5 cm2 only 40% healed baseline ulcer area with over 3 years duration, only 24% healed⁶. Additionally, patient 2 had a history of injury (burn) in the affected leg, and the skin bed was not optimum, surrounded by an atrophic skin, which could made the graft taking more difficult.

On the other hand, the encouraging results described with other composite skin substitute on leg ulcers (GraftskinTM)^{24,25} were only attained with more than one application of graft. It is possible that successive applications of the present composite skin substitute may bring better results.

CONCLUSION

In this pilot study there were no side effect upon the application of the graft, and it was demonstrated the feasibility of this skin substitute to treat refractory leg ulcers. We are now engaged in a randomized study with a more expressive number of cases to estimate the accuracy of such composite skin substitute on leg ulcers compared to compression.

RESUMO

Introdução: As úlceras de estase venosa de perna representam um desafio terapêutico associado a significativa morbidade e custo. Muitos avanços têm sido realizados no desenvolvimento e aplicação de substitutos de pele biológicos, mostrando que estes podem ser benéficos para pacientes com queimaduras, úlceras e outras lesões de pele. Os queratinócitos autólogos cultivados têm demonstrado utilidade no tratamento de úlceras de perna, apesar da ausência de componentes dérmicos poder comprometer seus efeitos e a qualidade da cicatriz. **Objetivo:** Avaliar o uso de um substituto composto da derme humana acelularizada e queratinócitos autólogos cultivados no tratamento de úlceras de perna. **Métodos:** Neste estudo piloto, dois pacientes com quatro úlceras crônicas de perna foram tratados com substituto de pele composto de derme humana acelularizada e queratinócitos durante um mês. **Resultados:** As quatro úlceras tiveram uma redução de tamanho de 72,21%, 36,36%, 60,93% e 15,90% em um mês de tratamento. Nenhuma reação adversa foi observada. A histologia e a imuno-histoquímica mostraram que a derme acelular humana estava livre de células do doador, e o substituto composto preservava a arquitetura normal de pele, incluindo componentes de membrana basal e epiderme estratificada. **Conclusões:** Os resultados preliminares sugerem que o substituto de pele composto pode ser uma alternativa para úlceras refratárias de perna.

Descritores: Úlcera Varicosa; Matriz Extracelular; Pele Artificial; Técnicas de Cultura de Tecidos; Técnicas de Cultura de Células.

REFERENCES

- 1. Miller F, Phillips TJ. Leg Ulcers. J Am Acad Dermatol. 2000;43:91-5.
- 2. Falanga V. Venous ulceration. J Dermatol Surg Oncol. 1993;19:764-71.
- 3. Phillips TJ, Dover JS. Leg ulcers. J Am Acad Dermatol. 1991;25:965-87.
- Valencia IC, Falabella A, Kirsner RS, Eaglstein WH. Chronic venous insufficiency and venous leg ulceration. J Am Acad Dermatol. 2001;44(3):401-21.
- Ghauri ASK, Taylor MC, Deacon JE, Whyman MR, Earnshaw JJ, Heather BP, Poskitt KR. Influence of a specialized leg ulcer service on management and outcome. Br J Surg. 2000;87:1048-56.
- Phillips TJ, Machado F, Trout R, Porter J, Olin J, Falanga V and the venous ulcer study group. Prognostic indicators in venous ulcers. J Am Acad Dermatol. 2000;43:627-30.
- 7. Phillips TJ, Kehinde O, Green H, Gilchrest B. Treatment of skin ulcers with cultured epidermal allografts. J Am Acad Dermatol.1989;21:191-9.
- Limat A, French LE, Blal L, Saurat JH, Hunziker T, Salomon D. Organotypic cultures of autologous hair follicle keratinocytes for the treatment of recurrent leg ulcers. J Am Acad Dermatol. 2003;48:207-14.
- Ramos-e-Silva M, Castro MCR. New dressings, including tissue-engineered living skin. Clinics in Dermatol. 2002;20:715-23.
- Jones L, Currie L, Martin R. A guide to biological skin substitutes. Br J Plastic Surg. 2002;55:185-93.
- Rennekampff HO, Kiessig V, Griffey S, Greenleaf G, Hansbrough JF. Acellular human dermis promotes cultured keratinocyte engraftment. J Burn Care Rehabil.1997;18:535-44.
- Omar AA, Mavor AID, Jones AM, Homer-Vanniasinkam S. Treatment of venous leg ulcers with dermagraft[®]. Eur J Endovasc Surg. 2004;27:666-72.
- Falanga V, Sabolinski M. A bilayered living skin construct (APLIGRAF) accelerates complete closure of hard-to-heal venous ulcers. Wound Repair Regen. 1999;7(4):201-7.
- Schonfeld WH, Villa KF, Fastenau JM, Mazonson PD, Falanga V. An economic assessment of Apligraf® (Graftskin) for the treatment of hard-to-heal venous leg ulcers. Wound Rep Reg. 2000;8:251-257.

- Weingarten MS. State-of-the-art treatment of chronic venous disease. Clinical Infectious Diseases. 2001;32:949-54.
- Sibbald RG, Torrance GW, Walker V, Attard C, MacNeil P. Cost-effectiveness of Apligraf in the treatment of venous ulcers. Ostomy Wound Manage. 2001;47(8):36-46.
- 17. Fivenson D, Scherschun L. Clinical and economic impact of Apligraf® for the treatment of nonhealing venous leg ulcers. Int J Dermatol. 2003;42:960-5.
- Medalie DA, Eming SA, Tompkins ML, Krueger GG, Morgan JR. Evaluation of human skin reconstituted from composite grafts of cultured keratinocytes and human acellular dermis transplanted to athymic mice. J Invest Dermatol. 1996;107:121-7.
- Yukna RA, Turner DW, Robinson LJ. Variable antigenicity of liophliized allogenic and lyophilized xenogenic skin in guinea pigs. J Periodontal Res. 1977;12:197-203.
- Green H et al. Growth of cultured humana epidermal cells into multiple epithelia suitable for grafting. Proc Natl Sci USA. 1979;765:5665-8.
- Ralston, R.D., Layton, C., Dalley, A.J., et al. Keratinocytes contract human dermal extracellular matrix and reduce soluble fibronectin production by fibroblasts in a skin composite model. Br J Plast Surg. 1997;50:408-15.
- Sheridan R, Choucair R, Donelan M, Lydon M, Petras L, Tompkins R. Acellular Allodermis in burn surgery: 1-year results of a pilot trial. J Burn Care Rehabil. 1998;19:528-30
- Ralston DR, Layton C, Dalley AJ, Boyce SG, Freedlander E, Mac Neil S. The requirement for basement membrane antigens in the production of human epidermal/ dermal composites in vitro. Br J Dermatol. 1999;140:605-15.
- Falanga V, Margolis D, Alvarez O, Auletta M, Maggiacomo F, Altman M, Jensen J, Sabolinski M, Hardin-Young J and The Human Skin Equivalent Investigators Group. Rapid healing of venous ulcers and lack of clinical rejection with an allogeneic cultured human skin equivalent. Arch Dermatol. 1998;134:293-300.
- Brem H, Balledux J, Sukkarieh T, Carson P, Falanga V. Healing of venous ulcers of long duration with a bilayered skin substitute: results from a general surgery and dermatology department. Dermatol Surg. 2001;27:915-9.